Three Terpenoids and a Tocopherol-Related Compound from Ricinus communis

by Qin-Gang Tan^a)^b), Xiang-Hai Cai^a), Zhi-Zhi Du^a), and Xiao-Dong Luo*^a)

a) State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, 132 Lanhei Road, Kunming 650204, P. R. China (phone: +86-871-5223177; fax: +86-871-5150227; e-mail: xdluo@mail.kib.ac.cn)
b) Guilin Medical College, Guilin 541004, P. R. China

Four new compounds named (3E,7Z,11E)-19-hydroxycasba-3,7,11-trien-5-one (1), 6α -hydroxy- 10β -methoxy- 7α , 8α -epoxy-5-oxocasbane-20,10-olide (2), 15α -hydroxylup-20(29)-en-3-one (3), and (2R,4aR,8aR)-3,4,4a,8a-tetrahydro-4a-hydroxy-2,6,7,8a-tetramethyl-2-(4,8,12-trimethyltridecyl)-2H-chromene-5,8-dione (4) were isolated from the MeOH extracts of the aerial parts of *Ricinus communis* L. by chromatographic methods. Their structures were elucidated by extensive spectroscopic experiments.

Introduction. – Ricinus communis L. (Euphorbiaceae), a herb or herbaceous shrub, is widely distributed in tropical regions and cultivated from tropical to extra tropical regions in the world [1]. This plant, which is called 'mahongliang' by the local Dai people in Yunnan province, P. R. China, is mainly used for the treatment of icteric hepatitis, arthritis, and constipation [2]. Recent researches have revealed the antifertility [3] and the inhibitory activity to HIV-1 reverse transcriptase [4] of the constituents from this plant. Previous chemical investigation on *Ricinus communis* L. led to the isolation of sterols [3], alkaloids [5][6], diterpenoids [7][8], coumarin, and flavonoids [5][9]. In order to find out the chemical basis of the favorable therapeutic effects, we have chemically investigated the aerial parts of Ricinus communis L., which led to the isolation of four new compounds named (3E,7Z,11E)-19-hydroxycasba-3,7,11-trien-5-one (1)¹), 6α -hydroxy- 10β -methoxy- 7α , 8α -epoxy-5-oxocasbane-20,10olide (2^1)), 15α -hydroxylup-20(29)-en-3-one (3^1)), and (2R,4aR,8aR)-3,4,4a,8a-tetrahydro-4a-hydroxy-2,6,7,8a-tetramethyl-2-(4,8,12-trimethyltridecyl)-2H-chromene-5,8dione (4) (Fig. 1). In this article, we report the isolation and structure elucidation of these new compounds.

Results and Discussion. – Compound **1** possesses a molecular formula of $C_{20}H_{30}O_2$, as evidenced by the HR-ESI-MS (m/z 325.2147, calc. 325.2143 for $C_{20}H_{30}NaO_2^+$, [M+Na]⁺), indicating six degrees of unsaturation. The IR spectrum showed absorption for OH (3426 cm⁻¹) and conjugated CO groups (1652 cm⁻¹), respectively. The presence of a conjugated CO group was supported by the UV absorption at 270 nm (log $\varepsilon = 4.74$) [10]. The ¹H-NMR spectrum of **1** (*Table 1*) displayed signals of three trisubstituted C=C bonds and four Me groups, two of them attached to sp²-C-atoms ($\delta(H)$ 1.85 (s,

¹⁾ Arbitrary numbering. For systematic names, see Exper. Part.

Fig. 1. Structures of the new compounds 1-4 from Ricinus communis L.

 $Me(18)^{1}$) and $\delta(H)$ 1.56 (s, Me(20))). With these functionalities, the two remaining degrees of unsaturation were ascribed to two ring systems. The chemical shifts at $\delta(C)$ 35.2, 27.6, 25.3, 15.8, and 28.8 in the ¹³C-NMR spectrum (*Table 1*) together with gemdimethyl at $\delta(H)$ 1.08 (s), 1.15 (s) in the ¹H-NMR spectrum indicated the presence of a cyclopropyl ring. These signals were typical for a casbane-type diterpenoid containing a 14-membered macrocyclic ring. By comparison the spectra data of 1 with those of (2E,6E,12E)-4-hydroxycasba-2,6,12-trien-5-one [11] and those of casbane diterpenoid [12], the differences consisted in that one of the Me groups in the latter two compounds was a CH₂ group in **1**. Considering that $\delta(H)$ 4.04 (d, J = 11.9) and 4.16 (d, J = 11.9)showed correlations with $\delta(C)$ 128.6 (*d*, C(7)), 139.3 (*s*, C(8)), and 35.4 (*t*, C(9)) in the HMBC spectrum (Fig. 2) of 1, the OH substitution was at C(19). The β -orientation of H-C(1) and H-C(2) was assigned on the basis of the close similarity of the coupling constant data with those of agrostistachin [13], of which the relative configurations were established by X-ray crystallographic analysis. This deduction was supported by the correlations of $\delta(H)$ 1.08 (s, Me(16)) with $\delta(H)$ 1.16 – 1.18 (m, H – C(1)) and 1.49 (t, J=9.0, H-C(2)) in the ROESY spectrum (Fig. 3) of 1. Thus, compound 1 was indentified as (3E,7Z,11E)-19-hydroxycasba-3,7,11-trien-5-one¹).

The molecular formula of compound **2** was determined to be $C_{21}H_{28}O_6$ based on the HR-ESI-MS (m/z 399.1788, calc. 399.1783 for $C_{21}H_{28}NaO_6^+$) and the DEPT data, suggesting eight degrees of unsaturation. The IR spectrum showed absorption for OH (3432 cm⁻¹) and conjugated CO groups (1649 cm⁻¹), respectively. The ¹H-NMR spectrum data of **2**¹) (*Table 1*) revealed the presence of five Me groups including one MeO group, two olefinic H-atoms, and two H-atoms attached to an O-bearing C-atom. The ¹³C-NMR and DEPT spectra data (*Table 1*) showed signals of two trisubstituted C=C bonds and two CO groups. The presence of a trisubstituted epoxide was deduced from the signals at $\delta(H)$ 2.70 (d, J=6.1) and $\delta(C)$ 63.5 (d) and 58.4 (s) [14]. Considering the disappearance of a C=C bond in the DEPT spectrum of **2** compared with that of **1**, the epoxidation occurred at C(7) and C(8). The ¹H- and ¹³C-NMR spectra of **2** were similar to those of hookerianolide A [15], except for the presence of

Table 1. ^{1}H -NMR (400 MHz) and ^{13}C -NMR (100 MHz) Data of **1** and **2** in CDCl₃. δ in ppm, J in Hz.

	11)		2 ¹)	
	$\delta(H)$	δ(C)	$\delta(H)$	δ(C)
H-C(1)	1.16-1.18 (m)	35.2 (d)	1.20-1.25 (m)	33.4 (d)
H-C(2)	1.49 (t, J = 9.0)	27.6(d)	1.48 (d, J = 8.4)	26.7(d)
H-C(3)	6.37 (d, J = 10.0)	143.2 (d)	6.56 (d, J = 7.7)	143.6(d)
C(4)		136.6 (s)		136.8 (s)
C(5)		199.6(s)		199.4 (s)
$CH_2(6)$ or $H-C(6)$	2.12-2.15, 1.96-2.00 (2m)	39.4 (t)	4.48 (t, J = 6.6)	72.4(d)
H-C(7)	5.08 (t, J = 8.0)	128.6(d)	2.70 (t, J = 6.1)	63.5(d)
C(8)		139.3(s)		58.4(s)
$CH_2(9)$	2.41-2.44, 1.78-1.81 (2m)	35.4(t)	2.78 (d, J = 15.0),	45.7(t)
			1.85 (d, J = 15.0)	
$CH_2(10)$ or $C(10)$	2.20-2.23, 2.06-2.09 (2m)	23.7(t)		106.8 (s)
H-C(11)	5.02 (t, J = 8.0)	119.7(d)	6.50(s)	143.0(d)
C(12)		136.9(s)		139.6 (s)
$CH_2(13)$	2.98-3.03, 3.49-3.53 (2m)	39.3 (t)	2.62-2.67, 2.14-2.19 (2m)	24.6 (t)
$CH_2(14)$	0.84 - 0.91, 2.10 - 2.15 (2m)	26.6(t)	2.12-2.16, 0.97-1.02 (2m)	22.5(t)
C(15)		25.3(s)		24.2 (s)
Me(16)	1.08(s)	15.8(q)	1.16(s)	15.7(q)
Me(17)	1.15(s)	28.8(q)	1.19(s)	28.2(q)
Me(18)	1.85(s)	11.6(q)	1.95(s)	13.0 (q)
$CH_2(19)$ or $Me(19)$	4.04 (d, J = 11.9),	59.7(t)	1.45(s)	19.5(q)
	4.16 (d, J = 11.9)			(1)
Me(20) or C(20)	1.56 (s)	15.9(q)		170.0(s)
MeO	.,	(1)	3.14 (s)	50.4 (q)

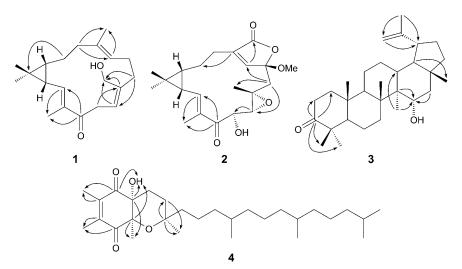


Fig. 2. Key HMBCs for compounds 1-4

Fig. 3. Key ROESY correlations for compounds 1-4

the ketone and the MeO groups, and the absence of an O-bearing CH group and the downfielded chemical shifts of the two olefinic C-atoms (C(3) and C(4)) in **2**. All these data suggested that the OH group in hookerianolide A at C(5) was replaced by an oxo group in **2**. From the correlations of δ (H) 6.50 (s, H–C(11)), 3.14 (s, MeO), 2.78 and 1.85 (2d, J = 15.0, CH₂(9)) with δ (C) 106.8 (s) in the HMBC spectrum (Fig. 2) of **2**, we assigned the MeO group to be placed at C(10). The β -orientations of H–C(6), H–C(7), and Me(19) were deduced from the ROESY spectrum (Fig. 3), in which the signal of δ (H) 1.48 (d, J = 8.4, H–C(2)) was correlated with δ (H) 1.20–1.25 (m, H–C(1)) and 4.48 (t, J = 6.6, H–C(6)), the signal of δ (H) 4.48 (t, t = 6.6, H–C(6)) with δ (H) 2.70 (t, t = 6.1, H–C(7)) and 1.45 (t (t Me(19)). Therefore, compound **2** was identified as $\delta \alpha$ -hydroxy-10 β -methoxy-7 α ,8 α -epoxy-5-oxocasbane-20,10-olide.

Compound 3 has the molecular formula of $C_{30}H_{48}O_2$ as evidenced by the HR-ESI-MS (m/z 463.3545, calc. 463.3552 for $C_{30}H_{48}NaO_2^+$), indicating seven degrees of unsaturation. The IR spectrum showed absorptions for OH (3434 cm⁻¹) and CO groups (1697 cm⁻¹), respectively. Analysis based on the combination of 1H -, ^{13}C -NMR and DEPT data ($Table\ 2$) revealed the presence of seven Me groups, a terminal C=C bond (δ (H) 4.58, 4.67 and δ (C) 109.7, 150.2), and an O-bearing CH group (δ (H) 4.15 (dd, J=4.9, 11.0) and δ (C) 69.6). The characteristic chemical shifts of the terminal C=C bond and five rings revealed that 3 was a lupane-type triterpenoid. The spectra data of 3 were similar to those of lup-20(29)-en-3 β ,15 α -diol [16], except that one of the OH groups was replaced by a ketone group. The downfielded shifts of C(2)¹) and C(4) in 3 indicated the location of the oxo group at the usual C(3) position. The only OH group was located at C(15), which was deduced from the correlations of δ (H) 4.15 (dd, J=4.9, 11.0) with δ (C) 47.9 (s, C(14)), 46.5 (t, C(16)), and 7.8 (t, C(27)) in the HMBC spectrum of 3 (t). The t-orientation of the OH group was deduced from the observation of the correlations of t(H) 4.15 (t) 4.15 (

a) Overlapped.

Table 2. ¹H-NMR (400 MHz) and ¹³C-NMR (100 MHz) Data of 3 and 4 in CDCl₃. δ in ppm, J in Hz.

	3 ¹)			4 ¹)	
	δ(H)	δ(C)		$\delta(H)$	δ(C)
CH ₂ (1)	1.37 – 1.41, 1.22 – 1.25 (2 <i>m</i>)	39.8 (t)	C(2)		87.0 (s)
$CH_2(2)$	2.41-2.44 (m)	34.1 (t)	$CH_2(3)$	1.87 - 1.92, 1.59 - 1.64 (2 <i>m</i>)	36.4 (t)
C(3)		218.2(s)	$CH_2(4)$	2.02-2.06, 1.68-1.73 (2m)	32.0(t)
C(4)		47.1(s)	C(5)		201.7(s)
H-C(5)	$1.28 - 1.31 \ (m)$	54.5 (d)	C(6)		141.9(s)
$CH_{2}(6)$	$1.51 - 1.53 \ (m)$	19.8 (t)	C(7)		146.9 (s)
$CH_{2}(7)$	1.86 - 1.89, 1.33 - 1.37 (2m)	36.9 (t)	C(8)		198.8 (s)
C(8)		42.2(s)	C(4a)		93.3 (s)
H-C(9)	$1.40 - 1.43 \ (m)$	50.2(d)	C(8a)		81.2 (s)
C(10)		37.0(s)	Me(1')	1.36 (s)	24.2(q)
$CH_2(11)$	1.48 - 1.51, 1.27 - 1.32 (2m)	21.5(t)	Me(2')	2.09(s)	13.4(q)
$CH_2(12)$	$1.63 - 1.67 \ (m)$	25.1(t)	Me(3')	2.10(s)	13.0(q)
H-C(13)	1.61 - 1.65 (m)	37.7(d)	Me(4')	1.39 (s)	25.8(q)
C(14)		47.9(s)	$CH_2(1'')$	1.55-1.63, 1.67-1.72 (2m)	41.4(t)
H-C(15)	4.15 (dd, J = 11.0, 4.9)	69.6 (d)	$CH_2(2'')$	1.40-1.43, 1.23-1.28 (2m)	22.3(t)
$CH_2(16)$	1.75 - 1.80, 1.28 - 1.34 (2m)	46.5 (t)	$CH_2(3'')$	1.05 - 1.24 (m)	37.3 (t)
C(17)		42.9(s)	H-C(4'')	$1.35 - 1.40 \ (m)$	32.7(d)
H-C(18)	$1.38 - 1.44 \ (m)$	47.9(d)	CH ₂ (5")	1.05 - 1.24 (m)	37.4(t)
H-C(19)	2.37-2.43 (m)	47.3(d)	$CH_2(6'')$	1.20-1.35 (m)	24.4(t)
C(20)		150.2(s)	$CH_2(7'')$	1.05-1.24 (m)	37.5 (t)
$CH_2(21)$	1.93 - 1.97, 1.31 - 1.36 (2m)	30.0(t)	H-C(8'')	1.35-1.39 (m)	32.8(d)
$CH_2(22)$	1.43 - 1.47, 1.17 - 1.23 (2m)	39.6 (t)	$CH_2(9'')$	1.05-1.24 (m)	37.5 (t)
Me(23)	1.05(s)	26.5(q)	$CH_2(10'')$	1.20-1.35 (m)	24.8(t)
Me(24)	1.01 (s)	20.9(q)	$CH_2(11'')$	1.10-1.15 (m)	39.3 (t)
Me(25)	0.93(s)	16.0 (q)	H-C(12'')	$1.48 - 1.54 \ (m)$	27.9(d)
Me(26)	1.15 (s)	16.2 (q)	Me(13")	$0.85 - 0.87^{a}$)	22.6(q)
Me(27)	0.96(s)	7.8(q)	Me(14")	$0.87 - 0.90^{a}$	19.7(q)
Me(28)	0.83(s)	19.0 (q)	Me(15")	$0.87 - 0.90^{a}$	19.8(q)
$CH_2(29)$	4.67 (s), 4.58 (s)	109.7(t)	Me(16")	$0.85 - 0.87^{a}$)	22.7(q)
Me(30)	1.67 (s)	19.3 (q)	ОН	3.86 (s)	

(s, Me(28)) and 1.15 (s, Me(26)) in the ROESY spectrum (Fig. 3). Therefore, **3** was deduced as 15α -hydroxylup-20(29)-en-3-one.

Compound **4** was deduced as $C_{29}H_{50}O_4$ by HR-ESI-MS analysis (m/z 485.3606, calc. 485.3606 for $C_{29}H_{50}NaO_4^+$). The DEPT spectrum data of **4** ($Table\ 2$) showed the signals of eight Me and eleven CH₂ groups, indicating the existence of a long aliphatic chain. Comprising the 1H - and ^{13}C -NMR data with those of VE-FPL (=7a-acetyl-3,4,4a,7-tetrahydro-4a-hydroxy-2,6,7-trimethyl-2-(4,8,12-trimethyltridecyl)cyclopenta[b]pyran-5(2H)-one) [17], together with the degrees of unsaturation, **4** was found to be a tocopherol-related compound. The ^{13}C -NMR and DEPT spectra data of **4** ($Table\ 2$) showed the presence of two conjugated CO groups and one tetrasubstituted C=C bond, which was supported by the strong absorption at 1679 cm $^{-1}$ and 253 nm in the IR and UV spectra of **4**, respectively. The close chemical shifts of the two CO groups (δ (C)

201.7, 198.8) and of the two olefinic C-atoms (δ (C) 146.9, 141.9) in **4** differentiated from those of in VE-FPL (δ (C) 205.0, 207.1 and 139.3, 163.1), indicating the left ring to be a six-membered in **4** instead of the five-membered in VE-FPL. This deduction was confirmed by the correlations of δ (H) 2.09 (s, Me(2')¹)) with δ (C) 201.7 (s, C(5)) and 141.9 (s, C(6)), of δ (H) 2.10 (s, Me(3')) with δ (C) 146.9 (s, C(7)) and 198.8 (s, C(8)) in the HMBC spectrum (Fig. 2) of **4**. Furthermore, the correlations of δ (H) (3.86, s, OH) with δ (C) 201.7 (s, C(5)), 93.3 (s, C(4a)), and 81.2 (s, C(8a)), of δ (H) 1.39 (s, Me(4')) with δ (C) 93.3 (s) and 81.2 (s), of δ (H) 1.36 (s, Me(1')) with δ (C) 87.0 (s, C(2)) and 36.4 (t, C(3)) were also observed. The relative configurations of the OH and the Me(4') groups were assigned as α , based on the observation of the correlations of δ (H) 3.86 (s, OH) with δ (H) 1.36 (s, Me(1')) and 1.39 (s, Me(4')) in the ROESY spectrum (Fig. 3). Thus, compound **4** was identified as (2R,4aR,8aR)-3,4,4a,8a-tetrahydro-4a-hydroxy-2,6,7,8a-tetramethyl-2-(4,8,12-trimethyltridecyl)-2H-chromene-5,8-dione.

Experimental Part

General. Column chromatography (CC): silica gel (SiO₂; Qingdao Marine Chemical Inc., P. R. China) and RP-18 (20–40 μ m, Merck). Optical rotations: Horiba SEPA-300 spectropolarimeter. UV Spectra: Shimadzu 210-A double-beam spectrophotometer; λ_{max} (log ε) in nm. IR Spectra: Bruker Tensor 27 spectrometer, KBr pellet and \tilde{v} in cm⁻¹. NMR Spectra: Bruker AM-500 spectrometer; δ in ppm with TMS as internal standard, J in Hz. HR-ESI-MS: VG Autospec-3000 spectrometer.

Plant Material. The aerial parts of *Ricinus communis* L. were collected in Kunming, Yunnan province, P. R. China, in March, 2007, and identified by Dr. *Chun-Xia Zeng*, Kunming Institute of Botany. A voucher specimen (NO. KUN20070310) has been deposited with the Herbarium of Kunming Institute of Botany, Chinese Academy of Sciences.

Extraction and Isolation. The air-dried aerial parts (16.0 kg) of Ricinus communis L. were crushed and extracted with 95% aq. MeOH (301×3) at r.t. (48 h × 3). After evaporation of MeOH, the viscous concentrate was dissolved in H_2O and partitioned with AcOEt (31×4) to afford AcOEt and H_2O extracts. The AcOEt extract (325 g) was subjected to SiO₂ CC (2.2 kg, 200–300 mesh) and eluted with CHCl₃/Me₂CO (1:0→1:1) to give eight fractions (Fr. I–VIII). Fr. III (63.0 g) was subjected to CC (SiO₂; 700 g) and eluted with petroleum ether (PE)/acetone (20:1→8:1) to afford five subfractions (Subfr. IIII – III5). Subfr. III3 (6.1 g) was chromatographed on RP-18 CC (MeOH/H₂O, 4:1→1:0), and then purified by SiO₂ CC (PE/AcOEt 12:1) to afford compound 4 (18.3 mg). Fr. V (103 g) was subjected to SiO₂ (1 kg) CC and eluted with PE/acetone (5:1→1:1) to afford six subfractions (Subfr. V1 – V6). Subfr. V2 (6.28 g) was subjected to RP-18 CC (MeOH/H₂O, 3:1→1:0), and then purified by SiO₂ CC (Pt/acetone 10:1) to afford compound 3 (34.8 mg). Separation of Subfr. V4 (4.5 g) with RP-18 CC (MeOH/H₂O, 4:1→1:0) to give compound 1 (102 mg). Subfr. V6 (7.8 g) was first subjected to RP-18 CC (MeOH/H₂O, 4:1→1:0) and then to SiO₂ CC (PE/AcOEt 8:1) to afford compound 2 (16.4 mg).

(3E,7Z,11E)-19-Hydroxycasba-3,7,11-trien-5-one (= rel-(1R,2E,10E,14S)-7-(Hydroxymethyl)-3,11,15,15-tetramethylbicyclo[12.1.0]pentadeca-2,6,10-trien-4-one; 1). Colorless oil. [α] $_{0}^{20}$ = - 16.8 (c = 0.72, CHCl $_{3}$). UV (CHCl $_{3}$): 270 (4.74), 236 (4.46), 202 (4.29). IR (KBr): 3426, 2926, 1652, 1452, 1274, 1066, 1003. 1 H- and 13 C-NMR (CHCl $_{3}$): Table 1. HR-ESI-MS: 325.2147 ([M + Na] $^{+}$, C $_{20}$ H $_{30}$ NaO $_{2}^{+}$; calc. 325.2143).

 6α -Hydroxy- 10β -methoxy- 7α ,8 α -epoxy-5-oxocasban-20,10-olide (= rel-(1R,5R,6R,8E,10S,12R)-6-Hydroxy-1-methoxy-3,8,11,11-tetramethyl-4,17-dioxatetracyclo[13.2.1.0^{3,5}.0^{10,12}]octadeca-8,15(18)-diene-7,16-dione; **2**). White amorphous powder. [α] $_0^2$ 0 = - 38.7 (c = 0.29, CHCl $_3$). UV (CHCl $_3$): 270 (4.78), 233 (4.54), 205 (4.45). IR (KBr): 3432, 1742, 1649, 1614, 1150, 960. 1 H- and 1 3C-NMR (CHCl $_3$): Table 1. HR-ESI-MS: 399.1788 ([M + Na] $^+$, C_{21} H $_{28}$ NaO $_6^+$; calc. 399.1783).

15α-Hydroxylup-20(29)-en-3-one (3). White amorphous powder. [α] $_{D}^{20}$ = +48.3 (c = 0.41, CHCl $_{3}$). IR (KBr): 3434, 2960, 2868, 1697, 1461, 1384. 1 H- and 13 C-NMR (CHCl $_{3}$): Table 2. HR-ESI-MS: 463.3545 ([M+Na] $_{+}$, C $_{30}$ H $_{48}$ NaO $_{2}$; calc. 463.3552).

 $\begin{array}{l} {\rm rel.}(2R,4aR,8aR)\text{-}3\text{,}4\text{,}4a,8a\text{-}Tetrahydro\text{-}4a\text{-}hydro\text{-}xy\text{-}2\text{,}6\text{,}7\text{,}8a\text{-}tetramethyl\text{-}2\text{-}(4,8,12\text{-}trimethyltridecyl)\text{-}} \\ 2H\text{-}chromene\text{-}5\text{,}8\text{-}dione\text{ (4)}. \text{ Colorless oil. } [a]_{0}^{20} = -12.7\text{ } (c=0.11, \text{CHCl}_3)\text{. UV (CHCl}_3)\text{: } 352\text{ } (3.05)\text{, } 339\text{ } (3.08)\text{, } 253\text{ } (4.66)\text{, } 228\text{ } (4.36)\text{, } 210\text{ } (4.24)\text{, } 201\text{ } (4.22)\text{. IR (KBr): } 3487\text{, } 2954\text{, } 1679\text{, } 1462\text{, } 1377\text{. } ^1\text{H-} \text{ and } ^1\text{C-NMR (CHCl}_3)\text{: } \textit{Table 2. HR-ESI-MS: } 485.3606\text{ } ([M+Na]^+, C_{29}H_{50}NaO_4^+; \text{ calc. } 485.3601\text{)}. \end{array}$

The authors are grateful to the *National Natural Science Foundation of China* (C30670214), *National Basic Research Program of China* (973 Program 2009CB522300), and the *Chinese Academy of Sciences* (KSCX2-YW-N-014, *XiBuZhiGuang* Project) for partial financial support.

REFERENCES

- [1] Editorial Committee of Flora of China of Chinese Academy of Sciences, 'Flora of China', Vol. 44, Science Press, Beijing, 1994, p. 88.
- [2] Yunnan Institute of Materia Medica, 'Illustrated Handbook for Medicinal Materials from Nature in Yunnan', Vol. 1, Yunnan Science and Technology Press, Kunming, 2005, p. 468.
- [3] X. Zhang, F. Han, P. Gao, D. Yu, S. Liu, Nat. Prod. Res. 2007, 21, 982.
- [4] H. X. Wang, T. B. Ng, Planta Med. 2001, 67, 669.
- [5] S. S. Kang, G. A. Cordell, D. D. Soejarto, H. H. S. Fong, J. Nat. Prod. 1985, 48, 155.
- [6] A. C. Leite, E. C. Cabral, D. A. P. dos Santos, J. B. Fernandes, P. C. Vieira, M. F. das G. F. da Silva, Ouim. Nova 2005, 28, 983.
- [7] L. Crombie, G. Kneen, G. Pattenden, J. Chem. Soc., Chem. Commun. 1976, 66.
- [8] D. Sitton, C. A. West, Phytochemistry 1975, 14, 1921.
- [9] S. M. Khafagy, Z. F. Mahmoud, N. A. E. Salam, Planta Med. 1979, 37, 191.
- [10] Y.-H. Choi, J. M. Pezzuto, A. D. Kinghorn, N. R. Farnsworth, J. Nat. Prod. 1988, 51, 110.
- [11] H. S. Santos, F. M. R. Mesquita, T. L. G. Lemos, F. J. Q. Monta, R. Braz-Filho, Quim. Nova 2008, 31, 601.
- [12] V. L. A. Moura, F. J. O. Monte, R. Braz Filho, J. Nat. Prod. 1990, 53, 1566.
- [13] Y.-H. Choi, J. Kim, J. M. Pezzuto, A. D. Kinghorn, N. R. Farnsworth, H. Letter, H. Wagner, Tetrahedron Lett. 1986, 27, 5795.
- [14] G. J. Greenland, B. F. Bowden, Aust. J. Chem. 1994, 47, 2013.
- [15] Y. Bai, Y.-P. Yang, Y. Ye, Tetrahedron Lett. 2006, 47, 6637.
- [16] R. Tanaka, K. Masuda, S. Matsunaga, Phytochemistry 1993, 32, 472.
- [17] J. Kitajima, K. Kimizuka, M. Arai, Y. Tanaka, Chem. Pharm. Bull. 1998, 46, 1647.

Received March 20, 2009